

Review

Open Access

Pulmonary bacterial pathogens in cystic fibrosis patients and antibiotic therapy: a tool for the health workers

Henrique Douglas M Coutinho*¹, Vivyanne S Falcão-Silva² and Gregório Fernandes Gonçalves²

Address: ¹Laboratório de Pesquisa em Produtos Naturais, Departamento de Ciências físicas e Biológicas, Centro de Ciências Biológicas e da Saúde, Universidade Regional do Cariri, Crato (CE), Brazil and ²Laboratorio de Genética de Microrganismos, Departamento de Biologia Molecular, Centro de Ciências Exatas e da Natureza, Universidade Federal da Paraíba, João Pessoa (PB), Brazil

Email: Henrique Douglas M Coutinho* - hdmcoutinho@gmail.com; Vivyanne S Falcão-Silva - vivyannefalcao@yahoo.com.br; Gregório Fernandes Gonçalves - gregorio_goncalves@yahoo.com.br

* Corresponding author

Published: 7 November 2008

Received: 27 May 2008

International Archives of Medicine 2008, 1:24 doi:10.1186/1755-7682-1-24

Accepted: 7 November 2008

This article is available from: <http://www.intarchmed.com/content/1/1/24>

© 2008 Coutinho et al; licensee BioMed Central Ltd.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract

Cystic fibrosis is the most common and best known genetic disease involving a defect in transepithelial Cl⁻ transport by mutations in the CF gene on chromosome 7, which codes for the cystic fibrosis transmembrane conductance regulator protein (CFTR). The most serious symptoms are observed in the lungs, augmenting the risk of bacterial infection. The objective of this review was to describe the bacterial pathogens colonizing patients with cystic fibrosis. A systematic search was conducted using the international bibliographic databanks SCIELO, HIGHWIRE, PUBMED, SCIRUS and LILACS to provide a useful and practical review for healthcare workers to make them aware of these microorganisms. Today, *B. cepacia*, *P. aeruginosa* and *S. aureus* are the most important infectious agents in cystic fibrosis patients. However, healthcare professionals must pay attention to emerging infectious agents in these patients, because they represent a potentially serious future problem. Therefore, these pathogens should be pointed out as a risk to these patients, and hospitals all over the world must be prepared to detect and combat these bacteria.

Introduction

Cystic fibrosis (CF) is the most common autosomal genetic disease in North America, affecting 1:2000 Caucasian individuals [1]. This disease is caused by mutations affecting the cystic fibrosis conductance regulator protein (CFTR) and is characterized by chronic lung malfunction, pancreatic insufficiency and high levels of chloride in sweat. Its high mortality index is evident when lung and spleen are affected, but other organs can also be affected. The persons affected die by progressive bronchiectasis and chronic respiratory insufficiency [2,3]. This disease affects persons without distinction of age or sex but can be

asymptomatic in a great number of cases [3]. Failure of innate defense mechanisms and the lack of mucociliary clearance in the airways stimulate primary and recurrent bacterial infections, blockage of airways, inflammation and chronic bacterial infections [4,5].

During the first decade of life of CF patients, *Staphylococcus aureus* and *Hemophilus influenzae* are the most common bacteria isolated from the sputum, but in the second and third decade of life, *Pseudomonas aeruginosa* is the prevalent bacteria. In Germany, analysis of the sputum from CF patients during a period of 12 months showed the pres-

ence of *P. aeruginosa* in 50% of these individuals, *S. aureus* in 63.3%, *Haemophilus influenzae* in 16.6%, *Stenotrophomonas maltophilia* in 15% and nontuberculous Mycobacteria (NTM) in 13.3% [6].

Due to this succession of bacterial populations in CF patients and due to the importance of these pathogens in prognosis, the objective of this article was to review and identify known and emerging bacterial pathogens associated with pulmonary problems and involved with cystic fibrosis. For this objective, a systematic search was conducted using the international bibliographic databanks SCIELO, HIGHWIRE, PUBMED, SCIRUS and LILACS. The uniterms Cystic fibrosis, infection and antibiotic therapy were used in a retrospective search between 1990 to 2007. Any articles with this theme, reporting bacterial pathogen associated with CF patients with no distinction of sex and age were selected and only the articles describing pathogens and the antibiotic therapy were really used.

Bacterial pathogens associated with pulmonary risk

Major pathogens

Mycobacterium sp

The nontuberculous Mycobacteria (NTM) are a group of microorganisms that is very common in chronic pulmonary diseases. The increase in the life expectancy of CF patients has also increased the prevalence of *Mycobacteria* in the CF population [7]. The clinical impact of these microorganisms in CF patients is unclear, because Oliver *et al.* [8] found that CF patients infected with NTM, observed for 15 months, did not show a decline in respiratory function. These microorganisms were isolated from older CF patients, all of them with perfect respiratory function, and were associated with a high frequency of *S. aureus* and a low one of *P. aeruginosa* when compared with patients without NTM, indicating that the presence of these bacteria may be taken as a good prognostic sign [9].

The most common NTM infecting CF patients are *Mycobacterium abscessus*, *Mycobacterium avium*, and *Mycobacterium intracellulare* [6], but Sermet-Gaudelus *et al.* [10] identified other NTM from CF patients, including *M. fortuitum*, *M. goodii* and *M. kansasii*. Today, the NTM more likely associated with the disease is *Mycobacterium abscessus* [11]. The identification of the causal species of NTM is essential and requires genetic techniques [12]. Treatment depends on the mycobacterial species. For *M. avium*, combined therapy with rifampicin, clarithromycin and ethambutol must be extended 12 months after negativation. *M. abscessus* infection is particularly resistant to therapy. Usual treatment is a one month course of intravenous imipenem or ceftazidime plus amikacin followed by oral clarithromycin plus ethambutol for at least 12 months after negativation. In case of local lesions, surgery is an option [12].

Staphylococcus aureus

Usually, this is the first pathogen to infect and colonize the airways of CF patients, being the most common pathogen [13]. This microorganism is prevalent in children and may cause epithelial damage, opening the way to the adherence of other pathogens such as *Pseudomonas aeruginosa* [14]. However, other studies indicate that *S. aureus* is a co-infective pathogen associated with *P. aeruginosa*. Together, the inflammatory process is more intense due the additive effect of these two pathogens [15]. Before the use of antibiotics in the treatment of infections, *S. aureus* was the causative agent of several deaths in children with CF. Today, this risk is not so serious, but CF patients not given the correct antibiotic therapy show a higher prevalence of *S. aureus* in the nasal epithelium when compared to treated patients [16]. About the prevalence of this pathogen, the same strain of *S. aureus* remains in the patient for 1–2 years [17].

Methicillin-resistant *S. aureus* (MRSA) has become a major nosocomial pathogen with a progressive increase in prevalence also in CF populations. The acquisition of MRSA occurred only in adulthood [18]. In Europe, the spread of MRSA varies widely among centers, ranging from 5 to 14% [19]. MRSA is a major pathogen in the hospital setting causing serious infections that usually present multiresistance to many antibiotics. Moreover, the increased frequency of this organism in the community, especially with carriage of virulence factors, including the presence of the virulence marker *pvl*, is a matter of concern [20,21].

Small colony variants (SCVs) of *S. aureus* constitute a bacterial population with distinctive phenotypic traits of *S. aureus* populations from CF patients [22]. These populations are involved with the colonization of older patients [23], but Sadowska *et al.* [24] isolated these strains from children between 1.5 and 9 years old with a SVR prevalence of 31.7%.

Pseudomonas aeruginosa

P. aeruginosa is an oxidase-positive Gram-negative motile rod [25]. Vonberg & Gastmeier [26] showed that this bacterium colonizes CF patients in more than 50% of cases. This bacterium is a part of the normal microbial population of the respiratory tract, where it is an opportunistic pathogen in CF patients. This is more prevalent in adult CF patients, as infection has been shown in 20% CF patients 0–2 years old while in 81% in adult groups (>18 years old) [27]. Aaron *et al.* [28] showed that all CF patients with chronic infections and older than 16 years are infected with *P. aeruginosa*, but Burns *et al.* [29] found that 97.5% of children had *P. aeruginosa*. The capacity of this bacterium to develop biofilm is a characteristic that

allows it to survive for very long periods in the lungs of CF patients [30].

Isolated from *P. aeruginosa* can be differentiated in terms of its morphotypes, including mucoid, not mucoid and those with biofilm, which vary their patterns of susceptibility to antibiotics. This differentiation causes several problems in the treatment, because is necessary identify the morphotype to choose the treatment strategy [31,32].

Burkholderia ssp

Burkholderia cepacia complex (BCC) is a complex of Gram-negative rod, aerobic, mesophilic and chemoorganotrophic [33]. This is a bacterial complex with nine genomic species (genomovars) [34,35]: genomovar I (*B. cepacia*), II (*B. multivorans*), III (*B. cenocepacia*), IV (*B. stabilis*), V (*B. vietnamiensis*), VI (*B. dolosa*), VII (*B. ambifaria*), VIII (*B. anthina*), IX (*B. pyrrocinia*) [35,36].

Infected CF patients show high levels of BCC in the salivary fluid, indicating the possibility of indirect transmission by kissing and sexual contact [36], but the transmission rates, prognosis and mortality are distinctly characteristic for each genomovar, as the treatment strategies [33,37]. Because the difficulties in the culture and identification of genomovar, this is one of the most important opportunistic bacterial pathogens of CF patients [38,39]. Other bacteria the same genus *Burkholderia*, as *Burkholderia gladioli* and *Burkholderia pseudomallei*, which are distinct from the *Burkholderia cepacia* complex have also been reported in patients with cystic fibrosis [40-42]. Members of *B. cepacia* complex are very resistant to antibiotic therapy because its genome is very plastic and suffers several mutations and adapts itself, making it a hard challenge for treatment. Its resistance is mainly due the production of enzymes with capacity to inactivate the substances used in the treatment [43]. By this fact., the accuracy and fast detection of this bacterium are essential to evaluate risks, prognostics and epidemiology of cystic fibrosis [35].

Minor pathogens

Achromobacter xylosoxidans

This bacterium is a Gram-negative rod, anaerobic, motile, oxidase and catalase positive and lactose non-fermentative. It is usually distributed in the environment, but can be a human pathogen causing bacteremia, meningitis and pneumonia [44]. This is a pathogen with a growing incidence in CF patients and a high coinfection rate with *P. aeruginosa* [45,46].

Inquilinus limosus

Coenye *et al.* [47] in 2002 isolated 8 strains from airway secretions of CF patients in the United States, that were identified as a new genus called *Inquilinus*, belonging to α -

proteobacteria and further identified as *I. limosus*. This bacterium is a mesophilic Gram-negative rod, non-spore forming. Due to its recent characterization, we have little knowledge about its natural habitat, prevalence and pathogenicity, but CF patients infected with this bacterium have been identified in hospitals in France, Spain and Germany [48,49].

Ralstonia sp

These bacteria are Gram-negative and non-fermentative rods, and little is known about the natural occurrence and the pathogenicity of bacteria from the genus *Ralstonia*, mainly due to their difficult identification, where they are usually misidentified as *P. fluorescens* or a member of the *Burkholderia cepacia* complex [50-54].

Reports indicate a low prevalence of pathogen from this genus in CF patients, but Coenye *et al.* [52] showed the permanence of this pathogen in the sputum of CF patients for more than 20 months.

Pandoraea apista

This is a Gram-negative and non-fermentative bacterium that over the years has shown a growing isolation frequency among CF patients, representing a possible emerging pathogen in these patients [55,56]. Atkinson *et al.* [57] analyzed sputum cultures from 2 adult CF patients (30 and 36 years old, respectively), and found both colonized by this bacteria and coinfecting with *P. aeruginosa*. This finding is very important due to the fact that these patients are first infected with *P. aeruginosa*, indicating that the latter pathogen may act as a starting point for *P. apista* infection.

Streptococcus pneumoniae

This microorganism is considered a transient pathogen in CF patients [58], mainly isolated from young CF patients [59]. The incidence is 5.5% in CF patients 12 and younger, but in children without the disease the frequency is 50% [60].

Stenotrophomonas maltophilia

This is a Gram-negative and non-fermentative rod that is frequently isolated from hospitals [61,62]. *S. maltophilia* is a pathogen of CF patients with a very constant incidence [63]. Goss *et al.* [62] observed that patients with *S. maltophilia* were older, showing a high rate of prior co-infection with *P. aeruginosa* and *B. cepacia*, but the prevalence of this pathogen in CF patients has been growing in the last years [64].

Haemophilus influenzae

This bacterium usually infects younger CF patients. In Brazil, 20.4% of CF children between 6 and 12 years old are infected by *H. influenzae* [65]. This bacterium undergoes

hyper-mutation, which can be related to its resistance to antibiotics, making treatment more difficult [66].

Bordetella bronchiseptica

This is a Gram-negative coccobacillus, non spore-forming, strictly anaerobic, and catalase and coagulase positive [67]. This bacterium is part of the microbiota of the upper respiratory tract of many animals [68]. Magalhães *et al.* [67], reported the presence of it in a 27-year-old CF patient associated with *S. aureus*, which can be a potential zoonotic infectious agent, aggravating the CF patient situation.

Treatment

Cystic fibrosis is characterized by chronic pulmonary infection with acute pulmonary exacerbation (APEs), where antibiotic therapy is necessary against opportunistic infections [69].

Previous studies have indicated that the presence of mucoidal *P. aeruginosa* was the most important risk factor for pulmonary deterioration [70,71]. By this fact, several articles indicating methods to control the colonizing pathogens in CF patients use *P. aeruginosa* as a microbial marker.

Gentamicin and tobramycin are recognized as standard antibiotics for the treatment of CF patients infected with *Pseudomonas aeruginosa*. Mulheran *et al.* [72] observed a higher utilization of gentamicin and tobramycin by pediatric patients and adults respectively. However, the authors make note of the greater cochleotoxic risk associated with gentamicin. Depending on the administration and dose used, tobramycin can be more or less efficient [73]. When this drug was used in a liposomal formulation and delivered as an aerosol, the drug bioavailability in pulmonary tissue and its effectiveness enhance [74,75].

Tests with animals have shown the augmentation of the amikacin concentration in the lung against *Pseudomonas aeruginosa*, when the drug is administered by ultrasonic nebulization or intravenously, but these levels decrease after the second administration [76].

Antibiotic combinations against *P. aeruginosa*, such as the use of polymyxins combined with a β -lactamic are useful in antipseudomonal therapy, as shown in the work of Dong & Chung-Dar [77].

Azithromycin displays interesting therapeutic results in the treatment of CF patients infected with *P. aeruginosa*. Wagner *et al.* [78] reported that azithromycin inhibits 80% of protein synthesis in *P. aeruginosa* PAO1, affecting bacterial growth and the expression/exportation of prod-

ucts that stimulates the immune system such as pyocyanin.

Other point of discussion is the objective of the treatment of *P. aeruginosa* infection: total eradication, using heavy doses of antibiotics with adverse symptoms, or the management of the infection, with a higher risk to develop the resistance? Few years ago, the eradication of chronic *P. aeruginosa* infection was considered impossible [79], but Ho *et al.* [80] e Pitt *et al.* [81] showed that new populations of *P. aeruginosa* (after eradication) were different of the first ones and more sensitive to the antibiotics, showing that persistent populations of *P. aeruginosa* in the airway would increase the antibiotic resistance with time because of prolonged exposure to antibiotics, as in the case of management, indicating the eradication as the most interesting strategy.

For other microorganisms such as *B. cepacia*, commonly resistant to several antimicrobial drugs used by CF patients, the better treatment choice is a drug combination. Combinations of two antibiotics from different classes such as meropenem-minocycline, meropenem-amikacin and meropenem-ceftazidime or three different antibiotics such as tobramycin, meropenem and an additional antibiotic were more effective than the use of any antibiotic alone [78]. Similar results were observed against *P. aeruginosa* by Dong *et al.* [77] who showed that the better treatment is the combination of meropenem/tobramycin or ceftazidime/tobramycin.

However, new therapeutic perspectives are needed, such as from the work of Zhang *et al.* [82] who evaluated the *in vitro* effectiveness of 150 antimicrobial peptides in multi-drug resistant strains of *P. aeruginosa*, *Stenotrophomonas maltophilia*, *Achromobacter xylosoxidans* and *S. aureus*. A better activity was observed for several peptides compared to most of the antibiotics used in the clinic. Similar results were obtained by Etienne *et al.* [83] who used defensins and observed a drastic reduction in bacterial growth.

The quality of life and life expectancy of CF patients have improved considerably as a result of the control of bronchopulmonary bacterial colonization and acute infectious exacerbations [82-85].

These reports indicate the necessity for more research into the discovery and rational design of new antibacterial drugs that will be more efficient in combating infections in cystic fibrosis patients. However, the use must be well defined. Our search indicates that the combination of 2 or more antibiotics may represent an interesting alternative in the CF treatment, colonized for any bacterial pathogen.

Other point of interest is the indication of aerosolized and biofilm-inhibitory drugs may control and avoid the colonization of the respiratory tract by several pathogens cited in this study. Maybe, using this several approaches, we will maximize the control of the colonizers and the infections that affect the CF patients.

Conclusion

Several factors affect transmission, such as the type of bacterial strain, the immune state of the patient and the use of contaminated medical equipment. Therefore, all CF patients infected or colonized the major pathogens cited in this article must be isolated in a single room because they represent sources for nosocomial transmission of the microorganism to other patients during the treatment [17,55].

Although the epidemiology of bacterial pathogens in CF patients has become more complex, the life expectancy of these patients continues to increase. This has led to a better control of the transmission of these pathogens by the separation of adults and children with CF in different treatment centers. Furthermore, the utilization of basic preventive guidelines (hand washing and use of masks, gloves and protectors), combined with disinfection techniques to be applied at home or hospital make control easier. These precautions help reduce the impact of infections in CF patients. In addition, educational programs to support administrative measures, guidelines for the control of nosocomial infections and the assistance to health-care workers and to the families of the patients to show the importance of these measures are essential tools for blocking the transmission of these bacterial pathogens to CF patients.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

VSFS and GFG contributed to conception and design, designed the review, carried out the literature research, and manuscript preparation. HDMC contributed to conception and design, carried out the manuscript editing and manuscript review. All authors read and approved the final manuscript.

References

1. Chu KK, Davidson DJ, Halsey TK, Chung JW, Speert DP: **Differential persistence among genomovars of the *Burkholderia cepacia* complex in a murine model of pulmonary infection.** *Infect Immun* 2002, **70**:2715-2720.
2. Goldman L, Bennett JC: **CECIL: Textbook of Medicine.** 21st edition. Guanabara Koogan, Rio de Janeiro; 2001.
3. Chaparro C, Maurer J, Gutierrez C, Krajden M, Chan C, Winton T, Keshavjee S, Scavuzzo M, Tullis E, Hutcheon M, Kesten S: **Infection with *Burkholderia cepacia* in cystic fibrosis: Outcome following lung transplantation.** *Am J Respir Crit Care Med* 2001, **163**:43-48.
4. Boucher RC: **New concepts of the pathogenesis of cystic fibrosis lung disease.** *Eur Respir J* 2004, **23**:146-158.
5. Accurso FJ: **Update in cystic fibrosis 2005.** *Am J Respir Crit Care Med* 2006, **173**:944-947.
6. Valenza G, Tappe D, Turnwald D, Frosch M, König C, Hebestreit H, Abele-Horn M: **Prevalence and antimicrobial susceptibility of microorganisms isolated from sputa of patients with cystic fibrosis.** *J Cyst Fibros* 2008, **7**(2):123-127.
7. Cullen AR, Cannon CL, Mark EJ, Colin AA: ***Mycobacterium abscessus* Infection in Cystic Fibrosis Colonization or Infection?** *Am J Respir Crit Care Med* 2000, **161**:641-645.
8. Oliver A, Maiz L, Canton R, Escobar H, Baquero F, Gómez-Mampaso E: **Nontuberculous mycobacteria in patients with cystic fibrosis.** *Clin Infect Dis* 2001, **32**:1298-1303.
9. Oliver KN, Weber DJ, Wallace RJ, Escobar H, Baquero F, Gómez-Mampaso E: **Nontuberculous mycobacteria I: multicenter prevalence study in cystic fibrosis.** *Am J Respir Crit Care Med* 2003, **167**:828-34.
10. Sermet-Gaudelus I, Le Bourgeois ML, Pierre-Audigier C, Offredo C, Guillemot D, Halley S, Akoua-Koffi C, Vincent V, Sivadon-Tardy V, Ferroni A, Berche P, Scheinmann P, Lenoir G, Gaillard JL: ***Mycobacterium abscessus* and Children with Cystic Fibrosis.** *Emerg Infect Dis* 2003, **9**:1587-1591.
11. Jönsson BE, Gilljam M, Lindblad A, Ridell M, Wold AE, Welinder-Olsson C: **Molecular Epidemiology of *Mycobacterium abscessus*, with Focus on Cystic Fibrosis.** *J Clin Microbiol* 2007, **45**:1497-1504.
12. Le Bourgeois M, Sermet-Gaudelus I, Catherinot E, Gaillard JL: **Mycobactéries atypiques et mucoviscidose.** *Archiv Pediatr* 2005, **12**(Suppl 2):S117-S121.
13. Saiman L, Siegel J: **Infection control in Cystic Fibrosis.** *Clin Microbiol Rev* 2004, **17**:57-71.
14. Lyczak JB, Cannon CL, Pier GB: **Lung Infections Associated with Cystic Fibrosis.** *Clin Microbiol Rev* 2002, **15**:94-222.
15. Sagel SD, Gibson RL, Emerson J, McNamara S, Burns JL, Wagener JS, Ramsey BV: **Impact of *Pseudomonas* and *Staphylococcus* Infection on Inflammation and Clinical Status in Young Children with Cystic Fibrosis.** *J Pediatr* in press. doi:10.1016/j.jpeds.2008.08.001.
16. Goerke C, Kraning K, Stern M, Döring G, Botzenhart K, Wolz C: **Molecular epidemiology of community-acquired *Staphylococcus aureus* in families with and without cystic fibrosis patients.** *J Infect Dis* 2000, **181**:984-989.
17. Branger C, Gardye C, Lambert-Zechovsky N: **Persistence of *Staphylococcus aureus* strains among cystic fibrosis patients over extended periods of time.** *J Med Microbiol* 1996, **45**:294-301.
18. Spicuzza L, Sciuto C, Vitaliti G, Di Dio G, Leonardi S, La Rosa M: **Emerging pathogens in cystic fibrosis: ten years of follow-up in a cohort of patients.** *Eur J Clin Microbiol Infect Dis* in press. DOI 10.1007/s10096-008-0605-4
19. Campana S, Taccetti G, Ravenni N, Masi I, Audino S, Sisi B, Repetto T, Döring G, Martino M: **Molecular epidemiology of *Pseudomonas aeruginosa*, *Burkholderia cepacia* complex and methicillin-resistant *Staphylococcus aureus* in a cystic fibrosis center.** *J Cyst Fibros* 2004, **3**:159-163.
20. Yang JA, Park DW, Sohn JW: **Novel PCR-restriction fragment length polymorphism analysis for rapid typing of staphylococcal cassette chromosome *mec* elements.** *J Clin Microbiol* 2006, **44**:236-238.
21. Tristan A, Bes M, Meugnier H: **Global distribution of Pantone-Valentine leukocidin-positive methicillin-resistant *Staphylococcus aureus*.** *Emerg Infect Dis* 2007, **13**:594-600.
22. Kahl B, Herrmann M, Everding AS, Koch HG, Becker K, Harms E, Proctor RA, Peters G: **Persistent infection with small colony variant strains of *Staphylococcus aureus* in patients with cystic fibrosis.** *J Infect Dis* 1998, **177**:1023-1029.
23. Vergison A, Denis O, Deplano A, Casimir G, Claeys G, DeBaets F, DeBoeck K, Douat N, Franckx H, Gigi J, Ieven M, Knoop C, Lebeque P, Lebrun F, Malfroot A, Pauquay F, Pierard D, Van Eldere J, Struelens MJ: **National survey of molecular epidemiology of *Staphylococcus aureus* colonization in Belgian cystic fibrosis patients.** *J Antimicrob Chemother* 2007, **59**:893-999.
24. Sadowska B, Bonar A, von Eiff C, Proctor RA, Chmiela M, Rudnicka W, Różalska B: **Characteristics of *Staphylococcus aureus*, isolated from airways of cystic fibrosis patients, and their small colony variants.** *FEMS Immunol Med Microbiol* 2002, **32**:191-197.

25. Hart CA, Winstanley C: **Persistent and aggressive bacteria in the lungs of cystic fibrosis children.** *Br Med Bull* 2002, **61**:81-96.
26. Vornberg RP, Gastmeier P: **Isolation of Infectious Cystic Fibrosis Patients: Results Of A Systematic Review.** *Infect Control Hosp Epidemiol* 2005, **26**:401-409.
27. Tramper-Stranders GA, Ent CK van der, Slieker MG, Terheggen-Lagro SW, Teding van Berkhout F, Kimpen JL, Wolfs TF: **Diagnostic value of serological tests against *Pseudomonas aeruginosa* in a large cystic fibrosis population.** *Thorax* 2006, **61**:689-693.
28. Aaron SD, Kottachchi D, Ferris WJ, Vandemheen KL, St Denis ML, Plouffe A, Doucette SP, Saginur R, Chan FT, Ramotar K: **Sputum versus bronchoscopy for diagnosis of *Pseudomonas aeruginosa* biofilms in cystic fibrosis.** *Eur Respir J* 2004, **24**:631-637.
29. Burns JL, Gibson RL, McNamara S, Yim D, Emerson J, Rosenfeld M, Hiatt P, McCoy K, Castile R, Smith AL, Ramsey BW: **Longitudinal assessment of *Pseudomonas aeruginosa* in young children with cystic fibrosis.** *J Infect Dis* 2002, **183**:444-452.
30. Costerton JW: **Cystic fibrosis pathogenesis and the role of biofilms in persistent infection.** *Trends Microbiol* 2001, **9**:50-52.
31. Martin DW, Schurr MJ, Mudd MH, Govan JR, Holloway BW, Deretic BW: **Mechanism of conversion to mucoid in *Pseudomonas aeruginosa* infecting cystic fibrosis patients.** *Proc Nat Acad Sci USA* 1993, **90**:8377-8381.
32. Saiman L, Mehar F, Niu WW, Neu HC, Shaw KJ, Miller G, Prince A: **Antibiotic susceptibility of multiply resistant *Pseudomonas aeruginosa* isolated from patients with cystic fibrosis, including candidates for transplantation.** *Clin Infect Dis* 1996, **23**:532-537.
33. Wallet F, Perez T, Armand S, Wallaert B, Courcol RJ: **Pneumonia Due to *Bordetella bronchiseptica* in a Cystic Fibrosis Patient: 16S rRNA Sequencing for Diagnosis Confirmation.** *J Clin Microbiol* 2002, **40**:2300-2301.
34. Ner Z, Ross LA, Horn MV, Keens TG, MacLaughlin EF, Starnes VA, Woo MS: ***Bordetella bronchiseptica* infection in pediatric lung transplant recipients.** *Pediatr Transplantation* 2003, **7**:413-417.
35. Melo-Coutinho HD: ***Burkholderia cepacia* complex: Virulence characteristics, importance and relationship with cystic fibrosis.** *Indian J Med Sci* 2007, **61**:422-429.
36. LiPuma JJ, Dulaney BJ, McMenamin JD, Whitby PW, Stull TL, Coenye T, Vandamme P: **Development of rRNA-based PCR assays for identification of *Burkholderia cepacia* complex isolates recovered from cystic fibrosis patients.** *J Clin Microbiol* 1999, **37**:3167-3170.
37. Soni R, Marks G, Henry DA, Robinson M, Moriarty C, Parsons S, Taylor P, Mahenthalingam E, Speert DP, Bye PT: **Effect of *Burkholderia cepacia* infection in the clinical course of patients with cystic fibrosis: A pilot study in a Sydney clinic.** *Respirology* 2002, **7**:241-245.
38. Vermis K, Coenye T, Lipuma JJ, Mahenthalingam E, Nelis HJ, Vandamme P: **Proposal to accommodate *Burkholderia cepacia* genomovar VI as *Burkholderia dolosa* sp. nov.** *Int J Syst Evol Microbiol* 2004, **54**:689-691.
39. Mahenthalingam E, Vandamme P, Campbell ME, Henry DA, Gravelle AM, Wong LT, Davidson AG, Wilcox PG, Nakielna B, Speert DP: **Infection with *Burkholderia cepacia* complex genomovars in patients with cystic fibrosis: Virulent transmissible strains of genomovar III can replace *Burkholderia multivorans*.** *Clin Infect Dis* 2001, **33**:1469-1475.
40. Kennedy MP, Coakley RD, Donaldson SH, Aris RM, Hohneker K, Wedd JP, Knowles MR, Gilligan PH, Yankaskas JR: ***Burkholderia gladioli*: Five year experience in a cystic fibrosis and lung transplantation center.** *J Cyst Fibros* 2007, **6**:267-273.
41. O'Carroll M, Kidd T, Coulter C, Smith H, Rose B, Harbour C, Bell S: ***Burkholderia pseudomallei*: another emerging pathogen in cystic fibrosis.** *Thorax* 2003, **58**:1087-1091.
42. Barth AL, Abreu e Silva FA, Hoffmann Vieira M, Zavascki P, Ferreira PAC, Cunha LG Jr., Albano RA, Marques EA: **Cystic Fibrosis patient with *Burkholderia pseudomallei* infection acquired in Brazil.** *J Clin Microbiol* 2007, **45**:4077-4080.
43. Miller MB, Gilligan PH: **Laboratory aspects of management of chronic pulmonary infections in patients with cystic fibrosis.** *J Clin Microbiol* 2003, **41**:4009-4015.
44. Liu L, Coenye T, Burns JL, Whitby PW, Stull TL, LiPuma JJ: **Ribosomal DNA-Directed PCR for Identification of *Achromobacter (Alcaligenes) xylosoxidans* Recovered from Sputum Samples from Cystic Fibrosis Patients.** *J Clin Microbiol* 2002, **40**:1210-1213.
45. Tan K, Conway SP, Brownlee KG, Etherington C, Peckham DG: ***Alcaligenes* infection in cystic fibrosis.** *Pediatr Pulmonol* 2002, **34**:101-104.
46. Van Daele S, Verhelst R, Claeys G, Verschraegen G, Franckx H, Van Simaey L, de Ganck C, De Baets F, Vaneechoutte M: **Shared Genotypes of *Achromobacter xylosoxidans* Strains Isolated from Patients at a Cystic Fibrosis Rehabilitation Center.** *J Clin Microbiol* 2005, **43**:2998-3002.
47. Coenye T, Goris J, Spilker T, Vandamme P, LiPuma JJ: **Characterization of Unusual Bacteria Isolated from Respiratory Secretions of Cystic Fibrosis Patients and Description of *Inquilinus limosus* gen. nov., sp. nov.** *J Clin Microbiol* 2002, **40**:2062-2069.
48. Chiron R, Marchandin H, Counil F, Jumas-Bilak E, Freydière AM, Bellon G, Husson MO, Turck D, Brémont F, Chabanon G, Segonds C: **Clinical and Microbiological Features of *Inquilinus* sp. Isolates from Five Patients with Cystic Fibrosis.** *J Clin Microbiol* 2005, **43**:3938-3943.
49. Wellinghausen N, Essig A, Sommerburg O: ***Inquilinus limosus* in patients with cystic fibrosis, Germany.** *Emerg Infect Dis* 2005, **11**:3390-3397.
50. Yabuuchi E, Kosako Y, Yano I, Hotta H, Nishiuchi Y: **Transfer of two *Burkholderia* and an *Alcaligenes* species to *Ralstonia* gen. nov.: proposal of *Ralstonia pickettii* (Ralston, Palleroni and Doudouroff 1973) comb. nov., *Ralstonia solanacearum* (Smith 1896) comb. nov. and *Ralstonia eutropha* (Davis 1969) comb. nov.** *Microbiol Immunol* 1995, **39**:897-904.
51. Coenye T, Vandamme P, Lipuma JJ: **Characterisation of unusual bacteria isolated from CF sputum.** *Pediatr Pulmonol* 2001, **22**:297.
52. Coenye T, Vandamme P, Lipuma JJ: **Infection by *Ralstonia* Species in Cystic Fibrosis Patients: Identification of *R. pickettii* and *R. mannitolilytica* by Polymerase Chain Reaction.** *Emerg Infect Dis* 2002, **8**:692-696.
53. Burns JL, Emerson J, Stapp JR, Yim DL, Krzewinski J, Loudon L, Ramsey BW, Clausen CR: **Microbiology of sputum from patients at cystic fibrosis centers in the United States.** *Clin Infect Dis* 1998, **27**:158-163.
54. Ferroni A, Sermet-Gaudelus I, Abachin E, Quesne G, Lenoir G, Berche P, Gaillard JL: **Use of 16S rRNA gene sequencing for identification of nonfermenting Gram-negative bacilli recovered from patients attending a single cystic fibrosis center.** *J Clin Microbiol* 2002, **40**:3793-3797.
55. Coenye T, Falsen E, Hoste B, Ohlén M, Goris J, Govan JR, Gillis M, Vandamme P: **Description of *Pandoraea* gen. nov. with *Pandoraea apista* sp. nov., *Pandoraea pulmonicola* sp. nov., *Pandoraea pnomenusa* sp. nov., *Pandoraea sputorum* sp. nov., and *Pandoraea norimbergensis* comb. nov.** *Int J Syst Evol Microbiol* 2000, **50**:887-99.
56. Jorgensen IM, Johansen HK, Frederiksen B, Pressler T, Hansen A, Vandamme P, Høiby N, Koch C: **Epidemic spread of *Pandoraea apista*, a new pathogen causing severe lung disease in cystic fibrosis patients.** *Pediatr Pulmonol* 2003, **36**:439-446.
57. Atkinson RM, Lipuma JJ, Rosenbluth DB, Dunne WM Jr: **Chronic Colonization with *Pandoraea apista* in Cystic Fibrosis Patients Determined by Repetitive-Element-Sequence PCR.** *J Clin Microbiol* 2006, **44**:833-8336.
58. Renders N, Verbrugh H, van Belkum A: **Dynamics of bacterial colonisation in the respiratory tract of patients with cystic fibrosis.** *Infect Genet Evol* 2001, **1**:29-39.
59. del Campo R, Morosini MI, de la Pedrosa EG, Fenoll A, Muñoz-Almagro C, Máz L, Baquero F, Cantón R: **Population Structure, Antimicrobial Resistance, and Mutation Frequencies of *Streptococcus pneumoniae* Isolates from Cystic Fibrosis Patients.** *J Clin Microbiol* 2005, **43**:2207-2214.
60. Muñoz C, Juncosa T, Gené A, Fortea J, Sécúli JL, Latorre C: **Microbiological study of the respiratory tract in children with cystic fibrosis.** *Enferm Infecc Microbiol Clin* 1996, **14**:142-144.
61. Denton M, Kerr KG: **Microbiological and clinical aspects of infection associated with *Stenotrophomonas maltophilia*.** *Clin Microbiol* 1998, **11**:57-80.
62. Goss CH, Otto K, Aitken ML, Rubenfeld GD: **Detecting *Stenotrophomonas maltophilia* does not reduce survival of patients with cystic fibrosis.** *Am J Respir Crit Care Med* 2002, **166**:356-361.

63. Goss CH, Mayer-Hamblett N, Aitken ML, Rubenfeld GD, Ramsey BW: **Association between *Stenotrophomonas maltophilia* and lung function in cystic fibrosis.** *Thorax* 2004, **59**:955-959.
64. Bauernfeind A, Emminger G, Horl G, Ott S, Przyklenk B, Weisslein-Pfister C: **Bacteriological effects of anti-*Pseudomonas aeruginosa* chemotherapy in cystic fibrosis.** *Infection* 1987, **15**:403-406.
65. Peltroche-Llacsahuanga H, Haase G, Kentrup H: **Persistent airway colonization with *Alcaligenes xylosoxidans* in two brothers with cystic fibrosis.** *Eur J Clin Microbiol Infect Dis* 1998, **17**:132-134.
66. De Baets F, Schelstraete P, Van Daele S, Haerynck F, Vanechoutte M: ***Achromobacter xylosoxidans* in cystic fibrosis: Prevalence and clinical relevance.** *J Cyst Fibros* 2007, **6**:75-78.
67. Magalhães M, Britto MCA, Bezerra PGM, Veras A: **Prevalence of potentially pathogenic bacteria in respiratory specimens of cystic fibrosis patients from Recife.** *J Bras Patol Med Lab* 2004, **40**:223-227.
68. Román F, Cantón R, Pérez-Vázquez M, Baquero F, Campos J: **Dynamics of Long-Term Colonization of Respiratory Tract by *Haemophilus influenzae* in Cystic Fibrosis Patients Shows a Marked Increase in Hypermutable Strains.** *J Clin Microbiol* 2004, **42**:1450-1459.
69. Ledson MJ, Gallagher MJ, Corkill JE, Hart CA, Walshaw MJ: **Cross infection between cystic fibrosis patients colonized with *Burkholderia cepacia*.** *Thorax* 1998, **53**:432-436.
70. Li Z, Kosorok MR, Farrell PM: **Longitudinal development of mucoid *Pseudomonas aeruginosa* infection and lung disease progression in children with cystic fibrosis.** *JAMA* 2005, **293**:581-588.
71. Parod RB, Gerard CJ, Zurakowski D, Nichols DP, Pier GB: **Pulmonary outcome in cystic fibrosis is influenced primarily by mucoid *Pseudomonas aeruginosa* infection and immune status and only modestly by genotype.** *Infect Immun* 1999, **67**:4744-4750.
72. Mulheran M, Degg C, Burr S, Morgan DW, Stableforth DE: **Occurrence and Risk of Cochleotoxicity in Cystic Fibrosis Patients Receiving Repeated High-Dose Aminoglycoside Therapy.** *Antimicrob Agents Chemother* 2001, **9**:2502-2509.
73. Coenye T, Lipuma JJ: **Multilocus restriction typing: A novel tool for studying global epidemiology of *Burkholderia cepacia* complex infection in cystic fibrosis.** *J Infect Dis* 2002, **185**:1454-1462.
74. Blumer JL, Saiman L, Konstan MW, Melnick D: **The Efficacy and Safety of Meropenem and Tobramycin vs Ceftazidime and Tobramycin in the Treatment of Acute Pulmonary Exacerbations in Patients With Cystic Fibrosis.** *Chest* 2005, **128**:2336-2346.
75. Burkhardt O, Lehmann C, Madabushi R, Kumar V, Derendorf H, Welte T: **Once-daily tobramycin in cystic fibrosis: better for clinical outcome than thrice-daily tobramycin but more resistance development?** *J Antimicrob Chemother* 2006, **58**:822-829.
76. Marier JF, Brazier JL, Lavigne J, Ducharme MP: **Liposomal tobramycin against pulmonary infections of *Pseudomonas aeruginosa*: a pharmacokinetic and efficacy study following single and multiple intratracheal administrations in rats.** *J Antimicrob Chemother* 2003, **52**:247-252.
77. Dong HK, Chung-Dar L: **Polyamines Increase Antibiotic Susceptibility in *Pseudomonas aeruginosa*.** *Antimicrob Agents Chemother* 2006, **50**:1623-1627.
78. Wagner T, Soong G, Sokol S, Saiman L, Prince A: **Effects of Azithromycin on Clinical Isolates of *Pseudomonas aeruginosa* From Cystic Fibrosis Patients.** *Chest* 2005, **128**:912-919.
79. Doring G, Conway SP, Heijerman HG, Hodson ME, Hoiby N, Smyth A: **Antibiotic therapy against *Pseudomonas aeruginosa* in cystic fibrosis: a European consensus.** *Eur Respir J* 2000, **16**:749-767.
80. Ho AS, Lee TWR, Denton M, Conway SP, Brownlee KG: **Regimens for eradicating early *Pseudomonas aeruginosa* infection in children do not promote antibiotic resistance in this organism.** *J Cyst Fibros* in press. doi:10.1016/j.jcf.2008.08.001
81. Pitt TL, Sparrow M, Warner M, Stefanidou M: **Survey of resistance of *Pseudomonas aeruginosa* from UK patients with cystic fibrosis to six commonly prescribed antimicrobial agents.** *Thorax* 2003, **58**:794-796.
82. Zhang L, Parente J, Harris SM: **Antimicrobial Peptide Therapeutics for Cystic Fibrosis.** *Antimicrob Agents Chemother* 2005, **49**:2921-2927.
83. Etienne O, Picart C, Taddei C, Haikel Y, Dimarcq JL, Schaaf P, Voegel JC, Ogier JA, Egles C: **Multilayer Polyelectrolyte Films Functionalized by Insertion of Defensin: a New Approach to Protection of Implants from Bacterial Colonization.** *Antimicrob Agents Chemother* 2004, **48**:3662-3669.
84. Ratjen F, Doring G: **Cystic fibrosis.** *Lancet* 2003, **361**:681-689.
85. Gibson RL, Burns JL, Ramsey BW: **Pathophysiology and management of pulmonary infections in cystic fibrosis.** *Am J Respir Crit Care Med* 2003, **168**:918-951.

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:
http://www.biomedcentral.com/info/publishing_adv.asp

